

SIOMICS V1.2 Manual

1. Prerequisites

In order to use this software, you should get the following things ready:

(1) You need to have python installed (Python 2 or Python 3).

You can download Python from [here](#).

Besides, you need Tkinter (Python 2) or tkinter (Python 3) module to enable the GUI.

For Windows users, the module was already included in the windows Python installer. ([link](#))

For Linux users, please see [here](#) for installation instructions.

(2) You would also have to install Java Runtime Environment.

You can check [here](#) for installation instructions.

If your java version is Java 6 or older, please upgrade it to Java 7 for avoiding any potential java Runtime Problems.

(3) You may also need to configure the Environment Variable for running java and python

A. For configuring Java Environment Variable, you can see [here](#) for instructions.

B. For configuring Python paths, you might see [here](#) for help.

2. Parameters

(1) Description of parameters

python SIOMICS.py

- -i <input_peak_sequences> Input TF ChIP-seq peak sequences should be in FASTA format, see [here](#).
Note: Please try to avoid including space or other special characters in the path of the input sequence.
- -o <output> This parameter used to specify the folder for output, all results will be put there.
If the specified output folder (e.g. c:\temp\output) already existed, the predicted results will be put on folder named output_1, output_2, etc (i.e. c:\temp\output_1, c:\temp\output_2, etc) to avoid mixing results for different runs together.
- -w <length_of_motifs> This parameter specifies the length of motifs, (Motif length range: 6-14).
- -m <maximal_number_of_output_motifs> The maximal number of motifs to be output.
- -s <support_value_of_motif_combination> The minimal number of sequences a motif module needs to occur in order to be considered as significant or frequent. For example, s=100 means the motifs co-occur at least 100 times to be claimed as frequent. If the instances of motifs in a module co-occur less than 100 times, this module will not be considered.
- -c <corrected_pvalue cutoff> The multiple comparison corrected p-value cutoff for motif module prediction.
- -r <number_of_iterations> The number of iterations.

(2) Recommended parameters for SIOMICS

- -w: 8, motif length =8
- -m: 100, in every iteration, 100 top motif candidates will be used to predict motif modules.
- -s: 1% of total number of sequences (Please keep in mind: s should be Integer, for example, the total input sequence 10,000, s=1%*10,000=100)
- -c: 0.01, corrected p-value cutoff 0.01
- -r: 20, 20 iterations at the maximal.

Note: If you can not get desired outputs under this recommended parameters, you might change those parameters based on your specific needs.

3. Software Usage

The following example shows you how to use the SIOMICS (both command line and GUI).

(1) Command line example

For example, if we want to identify motifs with length=8 from the provided "example_seq" dataset under the "example" directory. We can use the following command:

python SIOMICS.py -i example/example_seq -o example_output -w 8 -s 20 -c 0.01 -r 20 -m 100.

The meaning of the above parameters:

Try to identify motifs with length =8, corrected p-value < 0.01. The motifs need to co-occur at least 20 times to be claimed as modules. The maximal number of predicted motifs =100. The maximal iterations =20.

Note: The format of input sequence is the FASTA format.

If you do not want to specify every parameters by yourself, you can use the "batch_siomics.py" script we provided. This script can be used to run SIOMICS on a batch of peak sequences with default parameters.

Take the "example" folder included in the software as an example:

We can get the predictions for all datasets under the "example" folder by using the following command:

python batch_siomics.py example

SIOMICS will be run on each of sequence file under "example" folder sequentially.

The output files will be put into directory names as <DatasetName_out>

(2) GUI example

In order run GUI version of SIOMICS, just double click "SIOMICS_GUI.py". See the following GUI example:

Parameters		
Width of motifs (-w)	8	(e.g. 8 --motif with length 8)
Maximal number of output motifs (-m)	100	(e.g. 100 --consider top 100 motifs)
Significance level (-c)	0.01	(e.g. 0.01--corrected p-value cut 0.01)
Support value for pattern mining (-s)	20	(e.g. 1% (2000*1%=20),it need to be Integer--Motif combination co-occur at least 20 times(1%) to be discovered)
Maximal Iterations (-r)	20	(e.g. 20--The program will run 20 iterations at most)
Output (-o)	output	(e.g. output-- Results will be put under given folder 'output')

4. SIOMICS Results

When the software is running, you will see "Running..." shown in the bottom of the GUI. It might show "Not Responding" when SIOMICS is running on Windows, but it's OK. It will show "done" on the bottom of GUI once the results were obtained. SIOMICS will provide 5 files as the results of prediction in the output directory provided.

- (1) X.motifs
- (2) X.mc
- (3) X.motifs.JASPAR.pdf
- (4) X.motifs.TRANSFAC.pdf
- (5) X.mc.sif
- (6) X.tfbs
- (7) running.log

X.motifs is the result file of the predicted motifs in the format of frequency matrix

X.mc is the result file of motif modules predicted.

X.motifs.JASPAR.pdf is the comparison results between predicted X.motifs with JASPAR V.2010 database.

X.motifs.TRANSFAC.pdf is the comparison results between predicted X.motifs with TRANSFAC V.2010 database.

X.mc.sif is the interaction input for [cytoscape](#), ".sif" format was explained [here](#).

You can use X.mc.sif file to get the interaction network by cytoscape.

The instructions of how to load a simple interaction network (.sif) into cytoscape could be found [here](#).

If you are using SIOMICS_GUI version, the interaction network will be loaded into cytoscape by default.

X.tfbs is the TFBSs of the predicted motifs.

running.log records the commands and running time for SIOMICS software.

See the following example to see the meaning of X.motifs:

```
>M0      8.810747221152823      3.359771834161662E-5      8.0935338229
0.8764 0.0299 0.0518 0.0418
0.9081 0.0304 0.04 0.0215
0.8847 0.041 0.0416 0.0327
0.8378 0.064 0.0532 0.045
```

```

0.839 0.0654 0.0515 0.0441
0.8756 0.0456 0.0423 0.0365
0.9033 0.0333 0.0349 0.0286
0.8752 0.0337 0.0463 0.0449

```

The first line represents:

ID of motifs : M0

Scan_cutoff: 8.810747221152823 Used to define a putative TFBS of the motif.

lambda: 3.359771834161662E-5 Represent the probability of this motif occurring in random sequences (per nucleotide position).

MDScore: 8.0935338229 MDScore score used to represent the statistical significance of predicted motifs

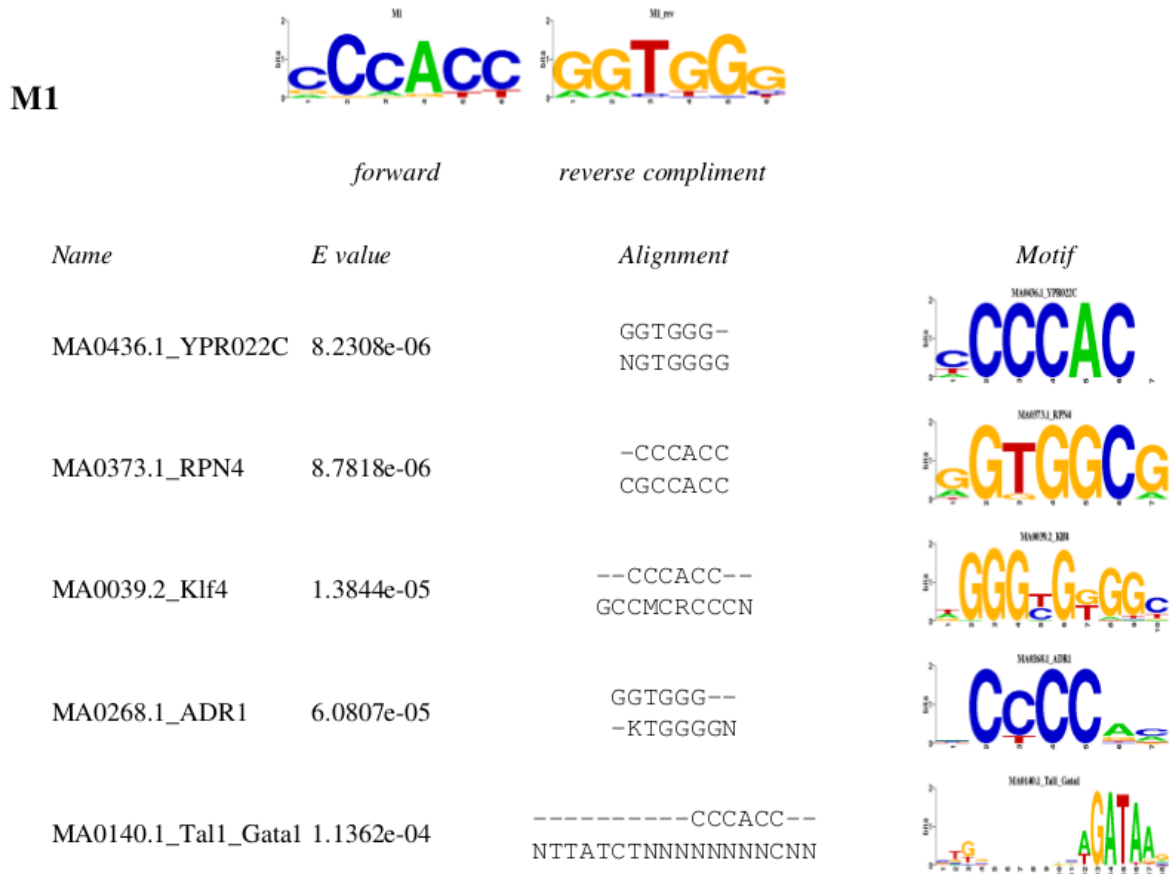
The remaining lines represent the frequencies of "A,C,G,T" in each position.

See the following example to see the meaning of X.mc:

```
M66 M21 (58) (6.7476668697e-10)
```

This denotes M66 and M21 were regarded as a motif module (co-occur in 58 sequences). The corrected pvalue is 6.7476668697e-10.

An example for X.motifs.JASPAR.pdf and X.motifs.TRANSFAC



Note: The above comparison between predicted motifs and known motif databases (TRANSFAC and JASPAR) were using [STAMP](#) with the default parameters. You could also compare the predicted motifs manually to other motif databases with different parameters by using [STAMP](#). You can simply use the predicted motifs as the "Input Motifs" of STAMP, then choose matched motif databases and parameters. For details about how to use STAMP, please refer to the [help](#).

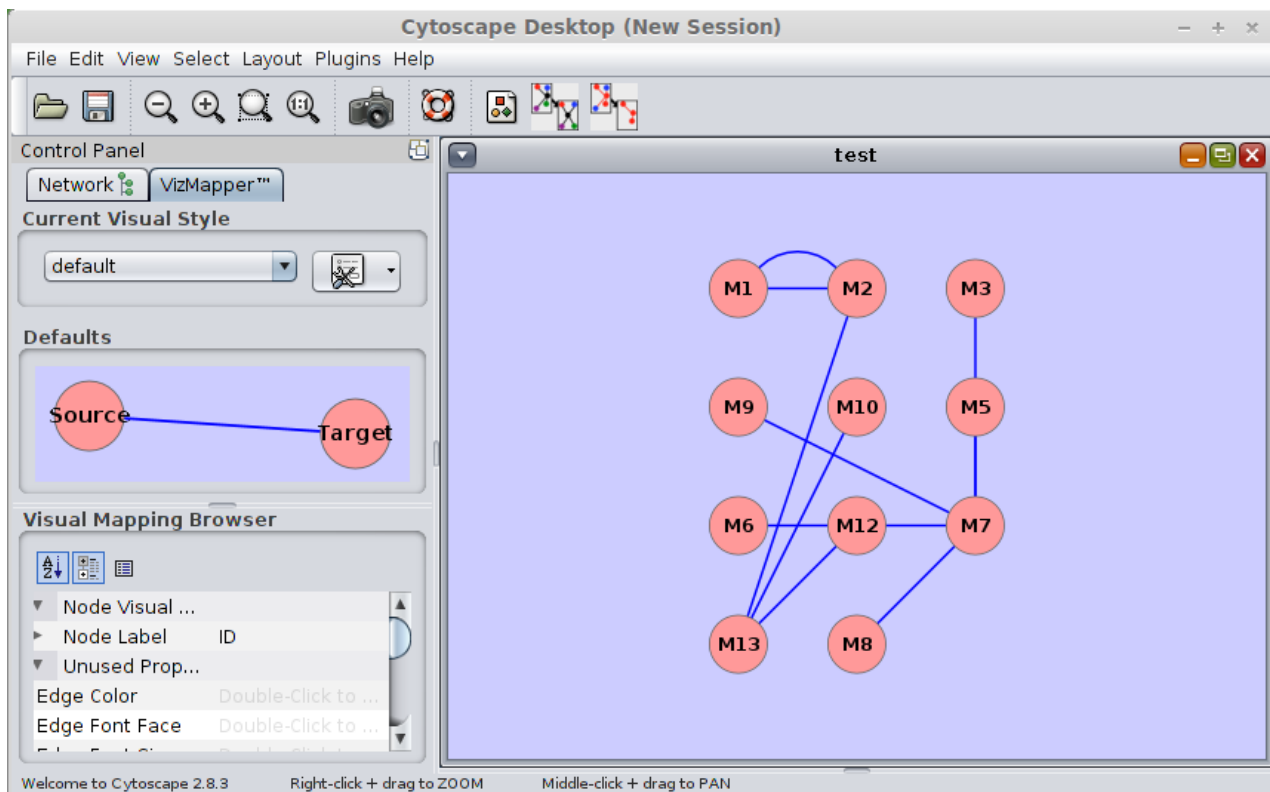
An example for X.mc.sif

```

M1 pp M2
M13 pp M12
M13 pp M10
M2 pp M1
M7 pp M9
M13 pp M2
M7 pp M6
M7 pp M5
M7 pp M8
M7 pp M3

```

pp means protein-protein interaction (M1= TF corresponds to motif M1, M2=TF corresponds to motif M2)
By using cytoscape, we will have the following interaction network:



Note: The cytoscape software will be automatically launched after the prediction was finished. If you do not like this, you could possibly use command Line version or the "batch_siomics.py" to get the predictions, which will skip this stage. After that, you might be able to use the ".sif" file generated to visualize the interaction network with cytoscape by yourself. You can get the details about how to use cytoscape from [here](#).

See the following example for X.tfbs

```
>M0:
M0,mm8_ct_UserTrack_3545_MACS_peak_19 range=chr1:13112912-13113786 5'pad=0 3'pad=0 strand=+ repeatMasking=N,609 GGGGGGGG
M0,mm8_ct_UserTrack_3545_MACS_peak_20 range=chr1:13645559-13646424 5'pad=0 3'pad=0 strand=+ repeatMasking=N,305 GGGTGGGG
M0,mm8_ct_UserTrack_3545_MACS_peak_32 range=chr1:36005234-36006169 5'pad=0 3'pad=0 strand=+ repeatMasking=N,42 GGGTGGGG
M0,mm8_ct_UserTrack_3545_MACS_peak_47 range=chr1:52897802-52898706 5'pad=0 3'pad=0 strand=+ repeatMasking=N,544 GGGGGGGG
M0,mm8_ct_UserTrack_3545_MACS_peak_72 range=chr1:72161911-72162847 5'pad=0 3'pad=0 strand=+ repeatMasking=N,356 GGGTGGGG
M0,mm8_ct_UserTrack_3545_MACS_peak_77 range=chr1:77336476-77337340 5'pad=0 3'pad=0 strand=+ repeatMasking=N,704 GGGTGGGG
```

This is the TFBSs information for predicted motif M0. Let's explain its format by using the first line:

```
M0,mm8_ct_UserTrack_3545_MACS_peak_19 range=chr1:13112912-13113786 5'pad=0 3'pad=0 strand=+ repeatMasking=N,609 GGGGGGGG
```

The above result represents M0 has a instance in peak (chr1:13112912-13113786,strand=+). The relative position of the instance is starting from 609 and the instance is GGGGGGGG.

The following is an example for running.log

```
Command using:
python SIOMICS.py
-i /home/jding/projects/SIOMICS_Aug/SIOMICS_Server_09_23/download/Software/SIOMICS_Linux/example/example_seq
-w 8
-m 100
-s 20
-c 0.01
-r 5
-o output
Running started from: 2013-09-29 19:58:48.735797
Running finished at: 2013-09-29 20:55:21.277166
```

You can retrieve the command you have used and time cost of the software from this running.log file

5. Contact Info

If you have any question regarding to the SIOMICS software or you have found any bugs, please feel free to contact us via xiaoman@mail.ucf.edu. For any non-academic use of this software, please also contact xiaoman@mail.ucf.edu.

